

Immuno-informatics analysis to identify novel vaccine candidates and design of a multi-epitope based vaccine candidate against Theileria parasites

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- 1 Title
- *Immuno-informatics* analysis to identify novel vaccine candidates and <u>design generation</u> of
 a multi-epitope based vaccine candidate against *Theileria annulataparasites*
- 4
- 5 Running title: Multi-epitope based vaccine candidates against *Theileria* annulata
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- 18 Key words: Theileria, immuno-informatics, multi-epitope vaccine, subunit vaccine
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20 Abstract:

Theileriosis poses a serious threat to ruminants in tropical and subtropical countries. It is a tick-21 22 borne disease, caused by an apicomplexan parasite, *Theileria*. The high disease burden in animals causes huge economic losses to marginal farmers. Further, with increasing cases of 23 resistance to commonly used drugs, it is highly desirable to develop better and cost-effective 24 vaccines against theileriosis. The high disease burden in animals lead to huge economic losses, 25 with increasing cases of resistance to commonly used drugs, there is a strong requisite for a cost-26 effective vaccine against theileriosis. The only available vaccine, live attenuated parasite 27 vaccine, has many drawbacks and hence is unsuitable for controlling this disease. IThe 28 immunoe-informatics has emerged as a useful tool in down selection of potential molecules for 29 30 vaccine development. In this study, we have usedused an immuno-informatics driven genomewide screening strategy to identify potential vaccine targets containing important and effective 31 32 dominant immunogens against *Theileria*. The proteome of *Theileria annulata* was screened for proteins with probability of plasma membrane localization or GPI anchor. The non-homologous 33 proteins non-homologous to the host (bovine) were selected and their antigenicity was analyzed. 34 The B-cell epitopes were identified in the selected proteins and were mapped in the modelled 35 structure of the proteins. A total of nineteen linear epitopes in twelve proteins, which are exposed 36 37 in the extracellular space and havinge the potential to induce protective antibodies were obtained. Additionally, CTL epitopes which are, peptides with 9-mer core sequence, were also identified, 38 modelled and docked with bovine MHC-I structures. The CTL epitopes showing high binding 39 energy with the bovine MHC-I were further engineered in silico to design a putative form a 40 multi-epitope vaccine candidate against *Theileria* parasites. The docking studies and molecular 41 dynamics studies with the predicted multi-epitope vaccine candidate and modelled bovine TLR-4 42

<u>exhibited showed</u>_strong binding energy, <u>which suggestings</u> that the complex is stable and <u>the</u>
<u>putative</u> multi-epitope vaccine candidate can be a potentially good candidate for vaccine
development.

46

47 Introduction:

48 *Theileria* spp. are tick--borne protozoan parasites which belong to the subphylum apicomplexa whichthat includes a number of other important pathogens such as Plasmodium, Babesia, 49 Toxoplasma, Cryptosporidium and Eimeria species. Theileria spp. are predominantly parasites of 50 51 ruminants and are transmitted transstadially to the host, transstadially, by ticks (Bishop et al., 2004; Sivakumar et al., 2014). These parasites cause huge economic loss to farmers in tropical 52 and subtropical countries. Among various species of these parasites, the two-most important ones 53 54 include T. parva and T. annulata, which cause economically important diseases in cattle, east coast fever and tropical theileriosis, respectively. T. parva is transmitted by Rhipicephalus 55 appendiculatus whereas T. annulata is transmitted by several species of Hyalomma. 56

57 The disease, theileriosis, is characterized by the lympho-proliferation of the host leucocytes after invasion by the parasites (Irvin, 1987). Other symptoms include fever, lymph node enlargement 58 59 and anaemia. The untreated animal usually dies within 3-4 weeks of the infection. This disease 60 kills over million animals every year and significantly reduces the productivity of the cattle (Norval et al., 1991; Gharbi et al., 2006). The cCurrent control measures for theileriosis include 61 the use of acaricides (for controlling vector), chemotherapy (drug such as buparavaquone) and 62 63 vaccination. Traditionally, control of tick infection has been performed by the application of acaricides. However, it is effective only when performed at the community level and regularly 64

for a prolonged period. The indiscriminate use of acaricides has been is leading to resistance in 65 tick, thus adding another dimension to the problem. The chemotherapy agents such as 66 parvaquone, buparvaquone and halofuginone are effective during the initial stages of infection 67 (McHardy et al., 1985). However, these chemotherapy agents are quite expensive and are not a 68 good choice for the marginal farmers. In addition, reports of development of resistance to these 69 70 chemotherapy agents areis a matter of deep concern (Mhadhbi et al., 2010). The only available vaccine for theileriosis is a live attenuated vaccine which is effective for providing protection to 71 vaccinated animal (Pipano, 1966). However, this vaccine needs a cold chain for transportation 72 73 and storage which leads to a huge increase in its cost (approximately \$8–12 per dose). Further, some vaccinated cattle, especially immunocompromised cattle, remain as a source of 74 transmission of parasites to other cattle (Maritim et al., 1989). 75

<u>TIn general, the subunit/multi-epitope based vaccines are generally -is-considered to be safer as</u>
compared to <u>a</u>_live attenuated vaccines (Kallerup, 2015). Also, the cost of production and
distribution of subunit/multi-epitope vaccine is <u>quite-comparatively</u> low, hence <u>theseit</u> would be
economical for the marginal farmers. Thus, developing <u>a</u> subunit and/or multi-epitope vaccine
for theileriosis, is <u>urgently required highly desirable</u>. However, <u>the</u> efforts to develop <u>the</u> subunit
vaccines against *T. annulata* and *T. parva* <u>till date</u> have not been successful <u>till date</u> (Nene and
Morrison, 2016).

LIt is possible to develop long_-lasting immunity against *Theileria* can be developed in cattle by
repeated challenge them_with *T. parva* and *T. annulata* sporozoites, in cattle by developing
neutralizing antibodies (Musoke et al., 1982). <u>TFurther, these neutralizing antibodies recognize</u>
the p67 protein of *T. parva* (Dobbelaere et al., 1984). <u>IThe immunization study</u> with recombinant
p67 antigen hass been been shown to induce immunity in 50% of vaccinated cattle (Musoke et

al., 2005). Also, in case of *T. annulata*, antibodies against parasite protein SPAG1 and Tams1 in *T. annulata* blocks the invasion of leucocytes by sporozoite (Williamson et al., 1989; Boulter et al., 1994; Boulter et al., 1998).

A large body of evidence indicates that cellular immunity also plays a major role in protection 91 against T. annulata and T. parva. The immunity can be adoptively transferred by CD8+, but not 92 CD4+ T cells from vaccinated animals. This strongly suggests that cytotoxic T lymphocytes 93 94 (CTL) play an important role in developing immunity against *Theileria* parasites (Taracha et al., 1995a; Taracha et al., 1995b). Further, major histocompatibility complex (MHC) class I-95 restricted CD8+ cells, the cytotoxic lymphocytes (CTLs), that target schizont-infected 96 lymphocytes also play a primary role in mediating immunity against *Theileria* in cattle 97 (MacHugh et al., 2009; Morrison, 2009). 98

The availability of comprehensive genomic, transcriptomic and proteomic datasets of Theileria 99 100 parasite have provided opportunities for *in silico* mining of novel candidates for vaccine design 101 (Gardner et al., 2005; Pain et al., 2005; Witschi et al., 2013). Immuno-informatics, which 102 integrates the transcriptomics and proteomics through advances in computational and molecular 103 immunological tools, has emerged as a new tool for identification of the target antigens for vaccine development (Davies et al., 2007; Tomar and De, 2014; Backert and Kohlbacher, 2015). 104 105 Since the experimental methods are difficult and time--consuming, the immuno-informatics 106 approach can narrow down a vast number of potential molecules to be tested, thus increasing the 107 chance of finding better candidates. Numerous studies have shown that epitopes based vaccines could be effective in elucidating protective immunity against various pathogens such as influenza 108 109 A, hepatitis B and, C virus, Leishmania and Shigella (Sominskaya et al., 2010; Stanekova and Vareckova, 2010; He et al., 2015; e Silva et al., 2016; Pahil et al., 2017). In this study, using 110

111 immunoe-informatics-driven vaccine target screening strategy, the available transcriptomic and 112 proteomic data for Theileria parasites were analyzed to predict proteins/epitopes for vaccine 113 development that could elicit protective humoral and cellular immune response. For an effective humoral response, the antigen must be antigenic and should possess B-cell epitopes, and for 114 115 effective CTL response, the peptide must be presented by MHC-I to the T cell receptor. Thus, 116 the model antigens for developing subunit/multi-epitope vaccine against *Theileria* should either 117 possess B-cell epitopes or be displayed by MHC-I to CTLs. UHere, using immuno-informatics, 118 we have identified 12twelve proteins containing 19nineteen epitopes which are likely to elicit the 119 humoral immune response. We have further designed -and constructed a multi-epitope subunit vaccine by combining CTL epitopes which have the potential to elicit the cellular immune 120 121 response against *Theileria* parasites (T. annulata and T. parva). The results provide a -a new 122 strategy for epitope--based vaccine development for Theileria parasites and indicate thathe immuno-informatics could beas an alternative strategy to rapidly discover potentialnew antigens 123 124 for subunit/multi-epitopes vaccine development against parasites.

125 Materials and Methods

126 A systematic workflow (Figure 1A) was generated for <u>the</u> identification of potential vaccine

127 candidates and construction of multi-epitope subunit vaccine against *<u>Theileria</u>* parasites (*T*.

128 *annulata* and *T. parva*) from the available transcriptomic/proteomic data (Gardner et al., 2005;

129 Pain et al., 2005; Witschi et al., 2013).

130 Sequence retrieval and sub-cellular localization

131 All protein sequences of *T. annulata* were retrieved from <u>the</u> National Centre for Biotechnology

132 Information (NCBI) database. The CD-HIT server was used to remove proteins having 90% or

more similarity. All the parameters recommended by <u>the</u> server were set to default (Huang et al.,
2010). The CELLO v.2.5 server was used for <u>the</u> identification of subcellular localization of all
the selected proteins. The plasma membrane proteins were selected with a confidence score of
>=2 (Yu et al., 2006). Further, all <u>the</u> proteins were analyzed for the presence of signal peptide
sequence <u>using SignaIP 4.1</u> and GPI anchor using <u>SignaIP 4.1 and</u> PredGPI server <u>respectively</u>,
with default settings (Pierleoni et al., 2008; Petersen et al., 2011).

139 Screening proteins non-homologous to <u>the</u> host and orthologous to *T. parva*

140 The selected plasma membrane proteins and proteins having both the signal sequence and GPI 141 anchor were further analyzed for screening the proteins which are non-homologous to the host, Bos taurus, using BLASTp server (Altschul et al., 1990) with a cut-off E-value 0.005. Proteins 142 143 predicted with E-value more than 0.005 were expected-likely to be non-homologous to Bos *taurus* and were selected for further analysis. Further, an orthologous search between T. 144 145 annulata and T. parva prediction was carried out using OrthoMCL tool to identify conserved 146 orthologous proteins (Chen et al., 2007). The query strategy was designed in such a way that the 147 average identity between two proteins should would be greater than 50%, and is-would be specific only to T. annulata and T. parva. 148

149 Antigenicity of proteins

150 The selected proteins (non-homologous to *Bos taurus* and orthologous to *T. parva*) from the

previous step were analyzed for the<u>ir</u> immunogenicity properties using Vaxijen - v2

152 (Doytchinova and Flower, 2007). The vaxijen is based on <u>an</u> alignment-free approach for antigen

prediction using auto cross_-covariance (ACC) transformation of protein sequences into uniform

154 vectors of principal amino acid properties. The classification by \underline{V} axijen is solely based on the

physicochemical properties of proteins. The threshold was kept to 0.5 to predict probable
antigenic and non-antigenic proteins.

157 **B-cell epitope prediction**

158 The <u>prediction of B-cell epitopes prediction for all the selected antigenic proteins from the</u>

159 previous step was performed using BCPREDS server with 90% specificity (El-Manzalawy et al.,

160 2008). BCPREDS server (B-cell epitope prediction) is based on Support Vector Machine (SVM)

161 classifiers and the training data-set containing 701 linear B-cell epitopes from Bcipep database

and randomly-701 random_non_-B-cell epitopes from SwissProt sequence database.

163 Further, the proteins were analyzed for the presence of putative transmembrane domains,

164 exposed and intracellular region(s) using TMHMM server

165 (http://www.cbs.dtu.dk/services/TMHMM/) (Krogh et al., 2001). Finally, those epitopes which

were exposed to <u>the extracellular surface</u> of the cell $\underline{s}_{\overline{s}}$ were selected for further analysis.

167 Model building, refinement and stereochemistry analysis of B-cell protein

168 The tertiary structure of the selected proteins which possess B-cell epitopes was built using RaptorX which is a web portal for protein structure and function prediction (Kallberg et al., 169 170 2012). RaptorX uses a threading--based modelling approach for model building. RaptorX server 171 predicts three--dimensional structures for protein sequences without close homologs in the 172 Protein Data Bank (PDB). RaptorX server can predict secondary and tertiary structures, contacts 173 solvent accessibility, disordered regions and binding sites. It can also assign some confidence scores to indicate the quality of a predicted 3D model such as p-value for the relative global 174 175 quality, GDT (global distance test) and uGDT (un-normalized GDT) for the absolute global

quality, and modelling error at each residue. The selected epitopes were then mapped on the
structure using <u>the</u> information obtained from the TMHMM prediction.

178 **<u>Prediction of</u>** T cell epitope(s) prediction and their specific MHC-I restriction element(s)

179 <u>The prediction of cytotoxic T-cell (CTL) epitopes was done using NetCTL 1.2 server (Larsen et</u>

180 al., 2007). Cytotoxic T-cell epitope (CTL) prediction was done using NetCTL 1.2 server (Larsen

181 et al., 2005). The threshold was set to 0.98 for high sensitivity and accuracy. All antigenic

182 proteins were analyzed for the presence of CTL epitopes. The CTL epitopes identified from the

HLArestrictor 1.1 (Erup Larsen et al., 2011). All seventy seven77 bovine MHC-1 alleles present

previous step were further screened for bovine MHC-I restriction element identification using

in the database were analyzed for their ability to bind to identified CTL epitopes and their

binding affinity with for respective MHC-I allele was also calculated. The Fthreshold for strong

binder (IC₅₀) was set to 50 and percentile rank was set to 0.5. <u>The T</u>threshold for weak binder

188 (IC₅₀) was set to 500 and percentile rank was set to 2.

183

189 Model building, refinement and stereochemistry analysis of bovine MHC-I

190 The sequence of bovine MHC-I alleles namely BoLA-3*05101, BoLA-1*00901, and BoLA-

191 1*00902 were downloaded from the Immuno Polymorphism Database (IPD) (Robinson et al.,

192 2013). Since crystal structures for these proteins were not present, a reliable model was built

using homology modelling. The best template for the homology modelling was obtained through

194 protein BLAST (pBLAST) against Protein Data Bank (PDB) and the three-dimensional model of

BoLA-MHC protein were built using modeller 9.19 (Webb, 2014). The generated model was

then refined by ModRefiner (Xu and Zhang, 2011) and stereochemistry analysis was done using

197 <u>PROCHECK, a protein parameters analysis tool PROCHECK</u> (Laskowski, 1993).

198 Tertiary model prediction for CTL epitopes

PEP-FOLD server was used to predict <u>the</u> three-dimensional structure of <u>alleach</u> CTL epitopes
 (Shen et al., 2014). Five probable structures were predicted by this server. The structure having
 the lowest energy model was selected as the final model for the <u>respective</u> epitope.

202 Interaction studies between selected MHC-1 allele and CTL epitope

203 The strong binders pairs of MHC-1 allele and CTL epitope obtained from HLA-restrictors were 204 then used for molecular docking studies using AutoDock vina (Trott and Olson, 2010). The hydrogen atoms were added to the selected MHC-I alleles and the file was then converted to 205 206 pdbqt format for docking studies with the CTL epitopes. For BoLA-3*05101, grid was set to -207 23.025, -15.521, -28.935 for X, Y and Z co-ordinates, respectively. The grid box for BoLA-1*00901 was set to 60.708, 61.642, 26.373 for X, Y and Z co-ordinates, respectively. In the case 208 209 of BoLA-1*00902 the grid box for X, Y and Z wasere set to 58.68, 64.412, 33.956 co-ordinates, respectively. The binding affinity of the ligands with its receptor was measured in kcal/mol. 210

Building and characterization of <u>the</u>multi-epitope subunit vaccine

All CTL epitopes selected after MHC-I restriction element analysis and interaction studies were fused together with the help of a linker AAY to build a multi-epitope vaccine molecule. The multi-epitope vaccine molecule thus obtained was analyzed for its non-homology to bovine (as described earlier). The antigenic properties of <u>the</u> multi-epitopes vaccine candidate was estimated by Vaxijen server (as described earlier). The allergenicity of the vaccine candidate was evaluated using AlgPred (http://crdd.osdd.net/raghava/algpred/)(Saha and Raghava, 2006). The physiochemical properties such as amino acid composition, theoretical pI, instability index etc., of the vaccine candidates were calculated by ProtParam (<u>http://web.expasy.org/protparam/</u>)
(Gasteiger, 2005).

221 Structure characterization of <u>the</u> multi-epitope vaccine candidate and bovine TLR-4

A template_-based tertiary structure prediction of the multi-epitope vaccine candidate was carried out by RaptorX server (Kallberg et al., 2012). <u>The Ff</u>urther, the refinement of modelled multiepitope vaccine candidate was done by using ModRefiner server as described previously (Xu and Zhang, 2011) and stereochemistry analysis was done using protein parameters analysis tool PROCHECK (Laskowski, 1993).

The sequence of bovine TLR-4 was downloaded from UniProt. Since crystal structure for this protein was not present, a reliable model was built using homology modelling. The best template for homology modelling was obtained through BLASTp against PDB database. Using a full length human TLR-4 as a template, <u>Aa</u> three-dimensional model of bovine TLR-4 protein was built <u>on modeller 9.19 using a full-length human TLR4 as a template</u>using modeller 9.19. The generated model was then refined by using ModRefiner and stereochemistry analysis was done using protein parameters analysis tool-PROCHECK.

234 Molecular docking of multi-epitope vaccine candidate with bovine TLR-4 receptor

The interaction studies between bovine TLR-4 and final subunit vaccine protein were carried out
with the help of a protein-protein docking server PatchDock (Schneidman-Duhovny et al., 2005).
The results were refined according to their binding score by FireDock server (Mashiach et al.,
2008).

239 Molecular dynamics simulation of bovine TLR-4 receptor with multi-epitope vaccine 240 candidate

241 To determine the stability of a protein-ligand stability-complex molecular dynamics (MD) simulation study of the complex is an important application tool. Molecular dynamics simulation 242 243 was carried out for the docked complex of the TLR-4 and multi-epitope vaccine protein for 100ns with a time integral of 2fs using Maestreo software. The simulation was done with Single 244 245 Point Charge (SPC) water as a solvent model and 0.15M-of NaCl buffer. The boundary 246 conditions wereas set to orthorhombic (a=b=c=10 Angstrom; $alpha=beta=gamma=90^{\circ}$). The 247 isothermal-isobaric pressure (NPT) ensemble weare set at temperature-300K (temperature) and 1 atm pressure. Nose-Hoover Thermostat method was used to maintain the temperature constant. 248 To examine the standard deviation and fluctuation of the backbone of the protein, the root mean 249 square deviation (RMSD) and root mean square fluctuation (RMSF) analysis were performed. 250

251 Results

Identification of plasma membrane and GPI anchored proteins of *Theileria* parasite. *annulata*

254 A systematic workflow based on the immuno-informatics approach was designed to identify the 255 potential vaccine candidates which would elicit an effective antibody response and to 256 designvelop a multi-epitope based vaccine candidate which would generate effective cell-257 mediated immune response against *T*-*heileria* parasites *annulata* (Figure 1A). The first step in 258 this workflow was to screen for T. annulata proteins which having the potential to be localized on the cell surface. For this tThe complete transcriptome/proteome of T. annulata comprising of 259 260 total 3783 protein sequences wasere downloaded from NCBI database for this purpose (Figure 261 1B). From all the protein sequences, tThe paralogous sequences with more than 90% similarity

262 were identified using CD-HIT server from all the protein sequences. Twenty-eight protein sequences were removed as paralogs while remaining 3755 protein sequences were selected for 263 264 further analysis. The selected 3755 protein sequences were submitted to the CELLO sub-cellular 265 localization servertool, to screen for the proteins predicted to be localized on the plasma membrane. CELLO is based on support vector machine (SVM) system to predict the probability 266 267 distribution for possible localization of the protein. The output of this software is in the form of different numerical values as gives different confidence scores for predicting the probability of 268 269 localization of the proteins in the cell. To get better stringency in screening of the plasma 270 membrane proteins, the confidence score of 2 or better was fixed. A total of 711 proteins obtained with the confidence score greater than 2 were selected for further analysis (Figure 1B). 271 Furthermore,, the criteria chosen for identification of proteins predicted to be localized on the 272 cell surface was that it should have a signal peptide along with a GPI anchor sequence. All the 273 274 3755 protein sequences were screened for the presence of a signal peptide sequence and a GPIanchorr sequence. From SignaIP analysis, out of Only 390 out of the 3755 proteins only 390 275 276 proteins were found to possess a signal peptide sequence based on Signal P 4.1 analysis. From PredGPI analysis, for GPI anchors signal, only 49 proteins containing a GPI-anchors signal 277 sequence were obtained. Only 24 out of these 3755 proteins were predicted to contain a signal 278 279 peptide sequence as well as a GPI-anchor-signal sequence (Figure 1B). 280 Ten out of these 24 proteins were also present in the plasma membrane analysis. After adding the 281 results from CELLO for proteins predicted to be localized at the plasma membrane-protein, and 282 proteins having both thea signal peptide sequence and athe GPI-anchor signal sequence, finally 283 726 protein sequences were finally selected for further analysis (Figure 1B).

Identification of <u>antigenic *T. annulata*</u> proteins non-homologous to *Bos taurus* and orthologous to *T. parva* and antigenic proteins of *T. annulata*

All 726 protein sequences from previous analysis based on <u>the</u> cell surface localization were analyzed for their non-homology to <u>the</u> host (*Bos taurus*) and orthology to *T. parva*. BLASTp analysis against *Bos taurus* resulted in 443 proteins with E-value above 0.005. It is assumed that these proteins are not present in the host (Figure 1B).

<u>The OrthoMCL server was usedemployed</u> for searching the orthologous or conserved protein
between *T. annulata* and *T. parva*. The criteria for orthologous protein selection was based on
conservancy of the protein in both the species with an average identity of greater than 50%. Out
of <u>the 443</u> selected proteins which are non-homologous to *Bos taurus*, only 32 proteins were
found to be conserved and only present <u>exclusively</u> in *T. annulata* and *T. parva*. These 32
proteins were selected for further screening (Figure 1B).

All 32 protein sequences selected from the previous step were submitted to Vaxijen web server to predict <u>the</u> antigenic properties of each protein. Vaxijen web server is an efficient tool for predicting antigenic properties of proteins with an accuracy of 70% to 89%. With the threshold of 0.5, oOut of the thirty two32 protein sequences, only twenty one21 protein sequences were predicted as probable potent antigen with the threshold of 0.5 (Table S1, S2). FOut of 21 protein sequences, fifteen out of 21 proteins sequences were found to be hypothetical.

302 Potential proteins of -*Theileria* parasites. *annulata* proteins for peptide/subunit vaccine
 303 development

B-cell epitopes are the antigenic determinants regions which are recognized by the specific
antibodies. The predicted antigenic proteins were then analyzed for <u>20 amino acid long</u> the

306 linear B-cell epitopes-of twenty residue long. BCPREDS server was used to predict the probable B-cell epitope and to minimize chances of false positive, the specificity was kept at 90% or more 307 308 to minimize chances of false positive. Four proteins, namely TA15480, TA21140, TA09756, and TA13665, were predicted not to possess any B-cell epitope with the defined cut off value (Table 309 S3). The B-cell epitope must be exposed to the extracellular regions of the cell to be recognized 310 311 by the antibodies., <u>hH</u>ence the <u>remaining 17</u> proteins with putative transmembrane domains were analyzed for the transmembrane, exposed and buried inside regions using TMHMM server. 312 313 Hence, rest 17 proteins were analyzed by TMHMM server (Table S4). The tertiary structures of 314 all these proteins were built using RaptorX, a web portal for protein structure and function prediction and the exposed epitopes were mapped on the structure (Figure 2, Figure S1). Five 315 316 proteins, namely TA15965, TA05820, TA14950, TA15795 and TA12045, were found to have no exposed B-cell epitopes, while the remainingst 12 proteins were found to contained B-cell 317 epitopes which were exposed to the cell surface (Figure 2, Figure S2). In these 12 proteins aA 318 319 total of nineteen-19 B-cell epitopes were predicted in these 12 proteins (Table 1). Presence of merozoite-piroplasm surface antigen (TA17050) which is one of the leading vaccine candidates 320 for the subunit vaccine against T. annulata (Gubbels et al., 2000) in our final list of 12 proteins, 321 322 validates authenticate the in silico analysis. TA17050 contains five B-cell epitopes with good 323 peptide score (more than 0.97 for all peptides) and an antigenic score of 0.5708. The remaining 324 <u>11 proteins also All other eleven proteins are potentially good targets for the development of</u> 325 subunit vaccine, especially TA09755 (FLPQTSRPSMGKKKGSFQLP-with peptide score 0.976) 326 showedexhibited the highest antigenic score (0.7931) among all eleven proteins. Tpr-related 327 protein family member (TA13385), which is unique to the Theileria parasite, could also be 328 considered as a good target for developing a subunit vaccine. Eight other peptides from the list

with scores more than 0.99, could be <u>evaluated</u> further tested _as <u>potential</u> peptide_-based vaccine
 <u>candidates</u> (Table 1).

Identification of cytotoxic T cell epitopes specific to bovine MHC-I restriction element(s)

332 The predicted 21 antigenic proteins were analyzed for the evtotoxic T cell (CTL) epitopes using 333 NetCTL server. A total of 179 CTL epitopes were predicted with a threshold value of 0.98 334 (Table S5). The CTL epitopes work in conjugation with the MHC-I present on the surface of antigen presenting cells. Hence, using HLA-restrictors server, all the 179 CTL epitopes were 335 336 analyzed for their ability to be recognized by MHC-I restriction elements of Bos taurus and to 337 predict the binding affinity of each CTL epitope with bovine MHC-I allele was predicted. Only 338 eleven-11 proteins containing twenty seven27 epitopes were predicted to have ashowed strong 339 binding affinity (IC₅₀ value <50 and percentile rank was also below 0.5) with MHC-I molecules (Table S6). Twenty-two Oout of these 2727 predicated T-cell epitopes, twenty two epitopes were 340 341 predicted to have ashowed strong binding affinity with BoLA-3*05101, fourteen-14 out of these 342 27 epitopes were predicted to have ashowed strong binding affinity with BoLA-1*00902, and 343 9nine out of these 27 epitopes were predicted to have ashowed strong binding affinity with BoLA-1*00901. Although many more MHC-I molecules were obtained which were predicted to 344 have ashowed strong binding to selected CTLs, but only these 3three MHC-I molecules (BoLA-345 346 3*05101, BoLA-1*00902, and BoLA-1*00901) were predicted to have ashowed strong binding 347 affinity for multiple epitopes (Table S6).

348 Molecular modelling and structure refinement of bovine MHC-I allele

The tertiary structure of BoLA-3*05101 was predicted by Modeller 9.19 with bovine MHC-I

molecule N*01801 (BoLA-A11) (3PWU) as the template. BoLA-3*05101 showed 84% identity

with N*01801. The root mean square deviation (RMSD) between <u>the</u> structure of modelled
BoLA-3*05101 and structure obtained after refinement was found to be 0.489<u>Å</u>. The predicted
tertiary structure of BoLA-3*05101 containsed 47.64% coils, 21.05% helixes and 31.30% beta
strands. The Ramachandran plot of this modelled protein obtained by PROCHECK tool
suggestsed that this was a highly reliable model (Figure S2).

356 Both BoLA-1*00902 and BoLA-1*00901 showed 76% and 86% identity, respectively with 357 cattle MHC-I N*01301 (2XFX'A') presenting an 11mer peptide from T. parva chain A. The RMSD value of modelled BoLA-1*00902 and BoLA-1*00901 protein, before and after 358 refinement, was calculated to be 2.682 Å and 1.713 Å respectively. The predicted structure of 359 BoLA-1*00902 contained 48.74% coils, 23.4% helixes and 27.86% beta strands while the 360 predicted structure of BoLA-1*00901 was found to have 45.54% coils, 24.02% helixes and 361 30.44% beta strands. The Ramachandran plot of modelled proteins obtained by PROCHECK 362 363 tool showed that the modelled proteins awere stable (Figure S2).

364 Modelled bovine MHC-I interacts strongly with identified CTL epitopes

365 For generating immune response iIt is necessary for the CTL epitopes to interact with the 366 respective MHC-I molecule with high binding affinity for generating specific immune response. In order to find the affinity between bovine MHC-I allele and identified CTL epitopepitope,e 367 molecular docking studies were performed using AutoDock Vina. All twenty seven27 CTL 368 epitopes modelled using PepFold server were docked with their respective MHC allele. The 369 370 predicted CTL epitopes "FVAWFYKLY" and "FLYKRDLPY" showed exhibited the highest binding affinity of -7.4 kcal/mol while "ILFTISLHY" exhibited showed the lowest binding 371 affinity of -5.5 kcal/mol with BoLA-N: 05101 MHC allele (Figure 3, Table 2). The predicted 372

373 CTL epitopes "FLYKRDLPY" showed highest binding affinity of -6.7 kcal/mol and

374 "IAFCIILYY" showed <u>the lowest binding affinity of -5.1 kcal/mol with BoLA-N: 00901 MHC</u>

allele. The predicte<u>d</u> CTL epitopes "IAFCIILYY" showed<u>displayed the</u> highest binding affinity

of -7.2 kcal/mol and "STIAMGLVY" showed<u>displayed</u> the lowest binding affinity of -4.4

377 kcal/mol with BoLA-N: 00902 MHC allele.

Further, TA12045 contains <u>the highest number of CTL epitopes which showed strong affinity to</u>
<u>18eighteen bovine MHC-I. Another protein, TA16125, contains 4four CTL epitopes which</u>
showed strong affinity to <u>12twelve bovine MHC-I (Table S6)</u>. Thus these proteins namely
TA15965, TA12045 and TA16125 can also be considered as good candidates <u>forof T cell</u>
mediated response against *T<u>heileria</u>*-*annulata*-parasite<u>s</u>.

383 <u>Design</u>Construction of multi-epitope subunit vaccine and prediction of allergenicity,
 384 physiochemical and antigenic properties

385 Based on the predicted binding affinity score of CTL epitope and bovine MHC-I restriction element analysis, all the 2727 CTL epitopes were fused together with a linker "AAY" to form 386 387 the multi-epitope subunit vaccine molecule. The final vaccine was composed of 321 amino acid 388 residues. The allergenicity of the vaccine construct was predicted using AlgPred against BLAST search on allergen representative peptides (ARPs). The result showed that the multi-epitopes 389 vaccine candidate would not act as is not an allergen. The molecular weight and theoretical pI of 390 391 this multi-epitope subunit vaccine candidate were calculated as 3.79 kDa and 6.75 respectively. 392 Based on the theoretical According to pI calculation this, the multi-epitope vaccine candidate 393 was found to be slightly acidic in nature. Further, the antigenicity of this potential vaccine

candidate construct was predicted to be 0.7286 as analyzed by Vaxijen server, suggesting that the
 multi epitope vaccine candidateit would beis antigenic in nature.

Homology modelling and model refinement of bovine TLR-4 and multi-epitope vaccine candidate

398 The three dimensional model of the multi-epitope vaccine candidate was generated using the 399 RaptorX server as no template was found against in the PDB database. Only one domain was predicted in the multi-epitope vaccine candidate by Tthe RaptorX server predicted only a single 400 401 domain for this multi-epitope vaccine candidate. The best template for the multi-epitope vaccine candidate was found to be 4AV3'A' with its p-value 6.61e⁻⁰⁵ (Figure 4A). The Ramachandran 402 plot of this protein revealed that 89.1% amino acids were located in the core region, 8.3% amino 403 404 acids were located in the allowed region, 1.6% amino acids were located in the generously allowed region and none was located in the disallowed region (Figure 4C). The predicted 405 406 secondary structure of the multi-epitope vaccine candidate was found to possess only helicxes 407 (Figure S3A).

As the crystal structure of bovine TLR-4 was not available, the tertiary structure of bovine TLR-409
4 was modelled using 4G8A (human TLR-4) as <u>a</u> template by Modeller 9.19 software (Figure 5A). Both <u>the</u> query coverage and identity w<u>eares</u> found to be 72%. The structure was then
refined using ModRefiner and validated with PROCHECK analysis. The Ramachandran plot of
<u>the</u> modelled bovine TLR-4 revealed that 79.6% amino acids were located in the core region,
17.4% amino acids were located in <u>the</u> allowed region, 1.8% amino acids were located in <u>the</u>

Multi-epitope vaccine candidate does not possesses bovine CTL and B-cell epitopes specific to *Theileria* parasites but not to the bovine host

The analysis for the presence of CTL epitopes in the multi-epitope vaccine protein by the 417 NetCTL server showed the presence of a total of forty nine49 CTL epitopes. Out of the forty 418 nine49 epitopes, twenty six26 epitopes were found to be the same as the previously predicted 419 420 epitopes in our analysis (Table S7). Remainingst twenty three 23 epitopes were the newly 421 predicted CTL epitopes. All the newly predicted CTL epitopes were subjected to BLASTp 422 against bovine for non-homology search and it was found that none of the these peptideepitopes were present in the bovine. Further, the discontinuous B-cell epitopes were also predicted from 423 424 the structure of the multi-epitope vaccine protein using EliPro server (Ponomarenko et al., 2008). A total of **9**nine conformational B-cell epitopes were predicted by the server (Figure 4B, Table 425 S8, Figure S3B). Similar to the CTL epitopes, all the predicted B-cell epitopes were also 426 427 analyzed for their -non-homology to the bovine and it was found that none of these B-cell 428 epitopes were present in the bovine proteome database. These results suggest that theis potential 429 multi-epitope vaccine protein is non-homologous to bovine and does not possess any predicted B-cell and CTL predicted epitopes from bovine. Hence, the putative multi-epitope vaccine 430 candidate would not generate an autoimmunity or tolerance in bovine. 431

432 Molecular docking of <u>the</u> multi-epitope vaccine candidate with bovine TLR-4 receptor

To identify predict the binding energy of this multi-epitope vaccine candidate with the bovine
TLR-4, docking of this multi-epitope vaccine candidate with TLR-4 was carried out using
PatchDock server which is based on the shape complementarity principle (Figure 5B).
PatchDock server generated 20twenty complexes which were scored according to the protein

437 surface, geometry and electrostatic complementarity. In order to refine and re-score the docking solutions of TLR 4 and multi-epitope vaccine candidate, tThe top ten complexes were then 438 submitted to the FireDock server in order to refine and re-score the docking solutions of TLR4 439 and multi-epitope vaccine candidate. The Ffinal docking model was chosen from the molecular 440 docking studies based on the binding score. The best structure of the multi-epitope vaccine 441 442 candidate and bovine TLR-4 showed a binding energy of -18.3 kcal/mol (Figure 5B). The attractive van der Waals force was calculated as -36.97 kcal/mol, the repulsive van der Waals 443 444 force was 49.96 kcal/mol, and the atomic contact energy was -2.10.

445 Molecular dynamics simulation of TLR-4 receptor with multi-epitope vaccine

The evaluation of the stability of the bovine TLR-4 and multi-epitope vaccine candidate complex 446 447 using molecular dynamics showed that the complex wais quite stable in SPC water type with 0.15M NaCl. The root mean square deviation (RMSD) of the backbone of the protein and the 448 root mean square fluctuation (RMSF) for all the side chain amino acid residues were analyzed 449 450 for a time period of 100ns. The interaction between the TLR-4 and the multi-epitope vaccine protein was found to be stable after 10ns (Figure 6A). The RMSF of side chain residues was 451 found to be in between $1^{A}_{A}^{\theta}$ to $3^{A}_{A}^{\theta}$ with a little variation (Figure 6B). A total of forty two42 452 hydrogen bonds were formed between the TLR-4 receptor and the multi-epitope vaccine protein. 453 454 The simulation analysis clearly shows that the multi-epitope vaccine protein can be recognized 455 by the TLR-4 and with form a stable complex.

456 Discussion:

457 <u>Various One of the few factors contributing to the of failure of the any antigen as a vaccine</u>
458 candidate under development stages are due to: (a) antigen not being exposed to immune

459 response and, (b) poor antigenic/immunogenicity response of antigen and, allergenicity to the host. Recently, with the advancement of *in-silico* analysis it has been possible to identify 460 461 molecule(s) which would likely possess maximum qualities y/properties to qualify asof a good antigen. The primary aim of this study was to identify the candidate gene(s) which havinge the 462 potential to induce an immune response against macroschizont stage of T. annulata. In this study 463 464 **W**e utilized a systematic immuno-informatics in this study approach to predict potential vaccine candidate(s) in *Theileria. annulata* parasites using the proteome databases of *T. annulata* 465 and *T. parva*, to designvelop a multi-epitope based vaccine. 466

One of the important criteria for selecting a potential antigen for vaccine development is that it 467 should be located on the surface of the cell so that it is more accessible for both the humoral and 468 the cellular immune system. The proteins are translocated to the cell surface using a signal 469 sequence for plasma membrane translocation or could be anchored to the 470 Glycosylphosphatidylinositol (GPI) moieties. These proteins which are exposed to the 471 472 extracellular surface are easily recognized and are likelyable to elicit an immune response when 473 used as the a target antigen for a vaccine development. PTherefore, the putative proteins from the 474 *Theileria* proteome database were thus examined for their translocation to plasma membrane using CELLO software. CELLO is based on support vector machine (SVM) system to predict 475 476 the probability distribution for possible localization of the protein win. Prediction of cellular

477 localization by CELLO is much<u>hich is</u> more reliable than <u>the</u> other sub-localization prediction

478 tool,-such as PSORTb-B. Furthermore, PSORTb program is trained only for the bacterial

479 <u>proteome (Gardy et al., 2005). We selected a confidence score of 2 or more for the sub</u>

480 <u>localization prediction based on our observation that in many cases the values lesser than 2</u>

481 resulted in predicting two different sub-localizations for the same protein. Proteins after

482 removing duplicates, were predicted for subcellular localization. A total of 711 proteins were predicted to be localized at the cell surface using the CELLO prediction server. To take into 483 484 account the proteins Further, the proteins that may also be present on at the cell surface using GPI moieties, . Wwe first looked for the proteins which havinge signal sequence peptide sequences 485 using the SignalP 4.1 server. Few research groups have also analysed the proteome of T. 486 487 annulata and T. parva for the identification of proteins containing signal peptide sequence and GPI anchor using the older version of SignalP (Pain et al., 2005; Weir et al., 2009; Woods et al., 488 489 2013; Bilgic et al., 2016). However, number of false positives in SignalP 4.1 is lesser compared 490 to the older versions (www.cbs.dtu.dk/services/SignalIP/performance.php). SignalP 4.1 uses a Hidden Markov Model (HMM) method for predicting the presence of signal peptide in the 491 492 protein. However, it does not predict the GPI anchors unlike the previous versions. PredGPI server was used to identify the GPI anchored proteins in our study. In order to identify those 493 494 proteins which may be GPI anchored, PredGPI server was used. The PredGPI uses a prediction 495 method which is coupled to HMM and SVM methods. PredGPI has been shown to outperform the other methods used for to-predicting GPI-anchors as it has a lower rate of false positive 496 predictions with respect to the other available methods (Pierleoni et al., 2008). Only 22twenty 497 498 two proteins were predicted to possess both the signal peptide and GPI anchor. The putative 499 plasma membrane and the GPI anchored proteins obtained from both the analysis were added for 500 further analysis.

In order to generate <u>an</u> immune response, <u>the</u> antigen must be recognized as non-self. Hence, the antigen for which homologous protein is present in <u>the</u> host is not <u>considered</u> a good candidate for vaccine development. Thus, we removed all the proteins homologous to *Bos taurus* <u>in our</u> study. Out of 733, only 443 out of the 733 proteins were predicted to beas non-homologous to Bos tauras. Further, an ideal vaccine candidate should be capable of stimulating an effective
immune response against various species of *Theileria*. Thus, to increase <u>the stringency</u>, <u>the query</u>
was set in such a way that only *T. annulata* or *T. parva* proteins arwould bee screened.

Antigenicity is one of the important features of an antigen for vaccine development. Vaxijen is the only server which uses alignment-independent prediction of protective antigens. It classifies antigen based on the physiochemical properties of the proteins. The prediction efficiency of this methods is found to be 70-80%. We finally found 21 Only twenty one proteins which were predicted as antigenic and also were found to be antigenic which qualified other screening conditions.

514 An effective vaccine should induce <u>a</u> specific immune response against specific

515 pathogens/antigen by selectively stimulating antigen-specific B-cells or CTLs or T helper cells. 516 In order to initiate the humoral response, antigens must be exposed and should be recognized by 517 the naive B-cells. Further, to initiate the cellular response, naive T cells need to be primed by the 518 peptide antigens bound to the major histocompatibility complexproteins (MHC) molecules on 519 the antigen presenting cells (APCs). The activation of CTLs (CD8+ cells) is a result of the 520 specific engagement of a 9 mer peptide to the MHC-I proteins. The CTLs eliminate the cells which they recognize as non-self, hence the immune system must be able to discriminate 521 522 between healthy and infected cells.

Thus, theo identify B-cell epitopes were predicted using the ,-BCPREDS server was used.
Further, B-cell epitopes must be accessible to the immune response. <u>H</u>, hence all the proteins
containing B-cell epitopes were modelled and <u>the epitopes were mapped on the structure</u>. Four
proteins did not show any B-cell epitopes and epitopes on <u>five 5 other proteins</u> were not

accessible to immune recognition. Finally, <u>11eleven</u> proteins were <u>found-predicted</u> to be good
targets for the development of subunit vaccine. One of the <u>11eleven</u> proteins was TA17050
which is one of the leading vaccine candidates, thus suggesting <u>the likeliness of the</u> other <u>10ten</u>
proteins <u>could alsoas be</u> good candidates for subunit vaccine development.

The CTL epitopes in all the antigenic proteins were identified using NetCTL server. NetCTL is a 531 532 quite reliable tool for the prediction of cytotoxic T lymphocytes and theits predictions done by 533 this is are based on proteasomal cleavage, Transporter associated with antigen presentation (TAP) 534 transport efficiency and MHC class I affinity. In comparison to the earlier version NetCTL1.0, the accuracy of prediction of cytotoxic T lymphocytes using NetCTL server is significantly 535 536 improved (Larsen et al., 2007). Although NetCTL is a tool designed for the prediction of human 537 CTL epitopes in a given protein, it can be used for bovine as well with equal efficiency. A total of 179one hundred and seventy nine CTL epitopes were identified using this prediction method. 538 539 Previously, studies conducted to identify CTL epitopes from T. parva specific CD8+ T cell line 540 by transfecting cells with parasite cDNA identified ten10 T. parva antigens having a total of 541 fifteen 15 CTL epitopes (Graham et al., 2006). It was initially intriguing to observe that Nnone of the antigens identified as CTL epitopes by this group were present in our analysis. However, 542 543 aAfter careful analysis, we found that the 10 proteins that this study identified did not possess 544 either a none of the proteins from which these CTL epitopes were identified, did not either 545 possess plasma membrane or signal sequence and GPI anchor together, sequence or they were not predicted to be localized to the plasma membrane. Using a similar strategy, three proteins 546 547 (TA15705, TA17545 and TA14970) were shown to be recognized by CD8 T cells from T. annulata infected cattle (MacHugh et al., 2011). TA14970 neither contains a signal sequence nor 548 a GPI anchor. Although, TA15705 (Ta9) and TA17545 possess the signal sequence, but these 549

550 proteins lack a GPI anchor. Hence, we did not include these proteins in our further analysis. The CTL epitopes must be presented by MHC-I on the surface of the antigen presenting cells to the 551 552 T-cell receptor for the activation of cell--mediated response. Hence, we predicted-calculated the binding energy of these CTL epitopes with all the bovine MHC-I molecules using HLA restrictor 553 554 server. The binding pairs which showed the highest energy were selected for further analysis. 555 Three MHC-I molecules, namely BoLA-3*05101, BoLA-1*00902 and BoLA-1*00901, were found to be the best, both in terms of binding energy and recognition to multiple CTL epitopes. 556 557 Hence these three molecules were further modelled to predict their three--dimensional structures. 558 Protein models having more than 90% of the residues in the core and allowed regions are generally considered as high--quality models. WHence, we found that the predicted model for 559 BoLA-3*05101, BoLA-1*00902, and BoLA-1*00901 were high-quality models based upon the 560 561 presence of more than 90% of their amino acid residues in the core and allowed regions. The docking of selected CTL epitopes with modelled BoLA-3*05101, BoLA-1*00902 and BoLA-562 563 1*00901 further confirmed the stability of the complex. Thus, twenty seven 27 CTL epitopes were found to be potential peptides for generating effective CTL response. Immunization with all 564 these twenty--seven, 9 mer peptides for the generation of CTL response would be a tedious 565 566 process., <u>hH</u>ence we <u>designed</u> a new protein containing all <u>the</u> CTL epitopes joined 567 with AAY linker. Immunization with a single protein (approximately 3.79 kDa) would be much 568 easier and cost effective if it could generate an effective cellular response. Thus, we re-analyzed 569 our newly designed again analyzed multi-epitope vaccine candidate for its ability to serve as an 570 ideal vaccine candidate. The newly designed engineered molecule was predicted found to be non-571 allergic while it but was antigenic in nature. The multi-epitope vaccine candidate was found to

possess <u>49</u>forty nine CTL epitopes and none of them w<u>ereas</u> found to be present in the bovine,
and twenty six <u>26</u> were specifically from *T<u>heileria</u>*. *annulata* parasites.-

574 Since the multi-epitope vaccine candidate is a novel protein, hence no template was found for 575 homologous modelling. Accordingly Thus, RaptorXAPTORX was used for generating a model for this protein. The possibility of the generated model being worse than the best of a set of 576 577 randomly generated models can be calculated by p-value. AThe lesser p-value represents the higher -quality model. For the alpha helix protein p-value should be lesser than 10⁻³-, and for the 578 beta--sheet protein p-value should be lesser than 10⁻⁴. The predicted structure of the multi-579 epitope subunit vaccine contained all alpha helicxes with a p-value of $6.61e^{-05}$ which indicate 580 581 that the structure predicted by RaptorX is a high--quality model. The un-normalized Global 582 Distance Test (uGDT) also estimate the model quality, like, if a protein contains greater than 100hundred residues and if the uGDT score is more than 50, it is a good indicator for high 583 quality model. The final multi-epitopes subunit vaccine candidate had the contained uGDT score 584 of 73. Forty-four percent of the protein residues wearre exposed in the subunit vaccine protein, 585 while and it is generally assumed was reported that if 15% of the protein residues are exposed 586 and solvent accessible, they can generate effective humoral and cellular immune response. 587

The modelled tertiary structure of <u>this</u> multi-epitope vaccine candidate suggests that the protein is quite stable and ordered. Further<u>more</u>, none of the discontinuous B-cell epitopes of the multiepitope vaccine protein were from bovine, suggesting that the new molecule would not generate any self-immune response to bovine.

The effective CTL response would be generated against the multi-epitope vaccine candidate
 would be generated only when it is presented to the CD8+ T cells by the antigen presenting cells.

The Toll-like receptors (TLRs) which are evolutionarily conserved proteins can sense foreign 594 molecules. They are characterized by an extracellular leucine--rich repeat domain and an 595 596 intracellular Toll/IL-1 receptor-like (TIR) domain. The TLRs play a fundamental role in the initiation of the immune response to the infectious agents through their recognition of the 597 conserved microbial molecular pattern. Out of all the TLRs, only TLR4 and TLR11 have been 598 599 shown to recognize the proteins (review (Takeuchi and Akira, 2010)). TLR-11 is absent in both 600 human and bovine, thus in our *in silico* analysis we tested the binding ability of multi-epitope 601 vaccine candidate with TLR-4 only. for which Wwe performed homologous modelling offor 602 bovine TLR-4 using human TLR-4 as a template. The binding energy of the designed multiepitope vaccine candidate and bovine TLR-4 was predicted found to be -18.3 kcal/mol and the 603 binding sites in the protein wereas distributed to the various regions of the proteins, suggesting 604 that the bovine TLR-4 would effectively recognize the vaccine candidate designed herein. Thus 605 606 overall in this study, we designed engineered a new vaccine candidate *in silico* containing 607 multiple epitopes of Theileia parasites. annulata.

608 **Conclusions**:

609 TIn conclusion, this study shows that the immuno-informatics driven genome-wide screening of vaccine targets for *Theileria* is a highly promising strategy to accelerate vaccine development for 610 this parasite. Based on this strategy, B-cell epitopes and CTLs were mapped from the proteome 611 612 of the *Theileria* parasite. This study provides a comprehensive analysis of *Theileria* proteins predicted to be better protective immunogens with high conservancy and, which would have the 613 potential for eliciting both neutralizing antibodies and T-cell responses. Thus, this study opens 614 615 new avenues for accelerating vaccine development by providing various potential molecules as novel vaccine candidates and a multi-epitope based vaccine candidate. 616

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- 781

782 Figure ligands:

- Figure 1. (A) Systematic workflow for identification of potential vaccine candidates and
- construction of multi-epitope subunit vaccine against *Theileria annulata* from the available
- 785 proteomic data. n, represent total number of proteins used for analysis.
- 786 ¹https://www.ncbi.nlm.nih.gov/;
- ²http://weizhong-lab.ucsd.edu/cdhit-web-server/cgi-bin/index.cgi?cmd=cd-hit;
- 788 ³http://cello.life.nctu.edu.tw/;
- ⁴http://gpcr.biocomp.unibo.it/predgpi/pred.htm;
- ⁵http://www.cbs.dtu.dk/services/SignalP/;

- ⁶ https://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE=Proteins 791
- ⁷ http://orthomcl.org/orthomcl/; 792
- 793 ⁸http://www.ddg-pharmfac.net/vaxijen/VaxiJen/VaxiJen.html;
- 794 ⁹http://ailab.ist.psu.edu/bcpred/;
- ¹⁰ http://raptorx.uchicago.edu/ 795
- ¹¹http://www.cbs.dtu.dk/services/NetCTL/; 796
- ¹²http://www.cbs.dtu.dk/services/HLArestrictor/; 797
- ¹³http://mobyle.rpbs.univ-paris-diderot.fr/cgi-bin/portal.py#forms::PEP-FOLD3; 798 3011a
- 799 ¹⁴Modeller9.19;
- ¹⁵AutodockVina: 800
- ¹⁶https://bioinfo3d.cs.tau.ac.il/PatchDock/; 801
- (B) Number of proteins obtained after various analysis. a. total proteins of *T. annulata*, b. 802
- proteins after removal of duplicates, c. plasma membrane proteins, d. proteins containing GPI 803
- anchor & signal peptide, e. proteins which are non-homologous to bovine, f. orthologous 804
- proteins specifically present in *T. annulata* and *T. parva*, g. antigenic proteins 805
- Figure 2. Molecular modelling of antigenic proteins of *Theileria annulata* containing B-cell 806
- epitope using RAPTOR X. (A) Ta 09755, (B) Ta 13065, (C) Ta 13810, (D) Ta 13385, and (E) Ta 807
- 16735. Identified epitopes mapped onto the modelled proteins are shown in red color, for buried 808
- epitopes, and blue color, for exposed epitopes using TMHMM. 809

Figure 3. Representative model of interaction between predicted CTL epitope through molecular
docking studies (A) FLYKRDLPY with BoLA-N:05101, binding energy -7.8 kcal/mol, (B)
FLYKRDLPY with BoLA-N:00902, binding energy = -7.7 kcal/mol, (C) WMVFFIVVY with
BoLA-N:00901, binding energy = -7.6 kcal/mol. Red represent epitope, blue represent modelled
bovine MHC-I and red represent CTL epitope.

Figure 4. Modelled structure of multi-epitope based subunit vaccine candidate: (A) Three
dimensional model of multi-epitope based subunit vaccine candidate obtained by homologous
modelling and refinement. The red color represents CTL epitopes and gray color represents AAY
linker, (B) B-cell epitopes of multi-epitope based subunit vaccine candidate represented in green
color, (C) Ramachandran plot analysis of the multi-epitope based subunit vaccine candidate
showing favored (91.7%), allowed (4.8%), generously allowed (2.6%) and disallowed (1%)
regions.

Figure 5. Docking studies of multi-epitope vaccine candidate and modelled bovine TLR-4: (A)
Three dimensional model of bovine TLR-4 obtained by homologous modelling and refinement,
(B) Docked complex of TLR-4 and multi-epitope based subunit vaccine candidate, (C) Binding
sites of multi-epitope vaccine candidate with bovine TLR-4. The interacting residues of the
putativeboth multi-epitope vaccine candidate and bovine TLR-4 are presented as surface
model.colored in green sphere.

Figure 6. Molecular dynamics simulation study of TLR-4 and multi epitope vaccine protein
complex. (A) Root Mean Square Deviation (RMSD) with a time duration of 100ns, grey line
represent side-chain and black line represent backbone of the protein chain (B) Root mean
Square Fluctuation (RMSF) of the docked complex side-chain with a time duration of 100ns.

- **Table 1.** Antigenic proteins with peptide score and antigenic score
- **Table 2.** Interaction studies with modelled bovine MHC-I with CTL epitopes
- 834 Supplementary figures
- Figure S1 (A) Molecular modelling of antigenic proteins of *Theileria annulata* containing B-cell
- epitope using RAPTOR X. The red color represent buried epitopes and blue represent exposed

using TMHMM, (B) Ramachandran plot analysis of modelled antigenic proteins of *T. annulata*

- 838 containing B-cell epitope using RAPTOR X.
- **Figure S2** Ramachandran plot analysis of bovine MHC-I alleles predicted by Modeller 9.19
- showing that all the predicted model are high quality model with more than 90% residue are in

core region. (A) BoLA-3*05101 (B) BoLA-1*00901 (C) BoLA-1*00902.

Figure S3 (A) Secondary structure of multi-epitope vaccine protein shows 90% residue are

helix, 9% residue are coil and no beta sheet are present in the structure (**B**) Conformational B-

845 vaccine protein was represented in red color with ribbon model. There are nine conformational

cell epitope in the multi-epitope vaccine protein predicted by EliPro server. The multi-epitope

- 846 B-cell epitope are present in the final multi-epitope subunit vaccine protein showed in different
- color with surface model.

844

- 848 Supplementary Table
- 849 **Table S1:** Annotation of antigenic proteins. (XLSX)

Table S2: Antigenic protein prediction using Vaxijen server with threshold 0.5. (XLSX)

Table S3: B-cell epitope prediction using BCPREDS with specificity 90%. (XLSX)

- Table S4: Analysis of epitopes present in cytoplasm, transmembrane and extracellular regions
 using TMHMM server. (XLSX)
- **Table S5**: T cell epitope prediction using NetCTL with a threshold of 0.98. (XLSX)
- **Table S6**: Identification of MHC-I restriction elements. (XLSX)
- **Table S7**: Identification of CTL epitopes generated in multi-epitope vaccine candidate. (XLSX)
- **Table S8**: Identification of B-cell epitopes generated in multi-epitope vaccine candidate.
- 858 (XLSX)



	S.	Gene ID	Position	Peptide sequence	Peptide	Antigenic
	No.				Score	Score
362	1	TA09755	132	FLPQTSRPSMGKKKGSFQLP	0.976	0.7931
	2	TA13065	157	KKPASDFEEQALEEYLKDKD	0.964	0.7595
363	3	TA13810	193	DEKEETSKKKYVLMVVVVV	0.972	0.7115
505	4	TA13385	336	AVGFPSITENWDSTAATGNG	0.993	0.6889
			375	EYKRHDPSKWPTDGMTTRTA	0.985	
864	5	TA16735	912	VNWEYVWEKEYKYYRMKMSN	0.986	0.6443
	6	TA17050	16	VISSVNAANEDEKKKEEKKD	1	
865			105	NLHPAQPKMFKKKGDKEYSE	1	
			212	FYTGDSRLKETYFELKDDKW	0.994	0.5708
			243	LNAMNSSWSTDYKPVVDKFS	0.991	
366			43	TSCENVTFKNVDSNTTELTV	0.978	
	7	TA11900	198	ELTTTKLVNVIVNGTQESIN	0.984	0.578
867	8	TA13820	24	EKNEQVTIDINKDATNPRKN	0.985	0.5741
	9	TA17055	175	HTVERDDESEEEAAITRVCC	0.999	0.5708
			39	LGYTLDTTIITSIGRDKINR	0.986	
868	10	TA16125	99	NVTEGGTSMYKYFVKVKGKW	0.963	0.5466
	11	TA21100	32	YNSTQNDTNTPRTGSYYNAV	0.999	0.5342
	12	TA12115	251	FINFICKIIPPGFPGQLLIQ	1	0.5083
869			37	LVKKTLFNRETDPSNNNFPF	0.963	
370						
371						
372						
372 373						
872 873 874						
 371 372 373 374 375 376 						

Table 1: Antigenic proteins with peptide score and antigenic score

879 Interaction	n studies with BoLA-N	:05101 and
	CTL epitopes	
880 MHC-I	CTL epitopes	Binding energy
		(Kcal/mol)
881	LYEYVLYLY	-5.8
	FVAWFYKLY	-7.4
382	FSNLYSGYY	-6.7
102	FLYKRDLPY	-7.4
	IAFCIILYY	-6.9
83	FINLTLLTY	-6.4
	ITDVLIYIY	-6.8
84	ILLTFNHLY	-7.1
04	ILFTISLHY	-5.5
	CSLYFVVLY	-6.3
BoLA-N:05	101 KMGHLTIYY	-6.4
BOLA-N.05	STIAMGLVY	-6.5
36	SSFNILLSY	-6.4
	FINLVHYYY	-6.2
	FTEHNSLEY	-6.6
87	YSLLFFYLY	-5.8
	SLLFFYLYY	-6.9
88	FALDIMTKY	-5.7
	YIKEYFSLY	-7.1
	VVFDYSVKY	-5.5
89	VFSIVSSLY	-6.5
	FFELLPSLY	-6.7

Table 2: Interaction studies with modelled bovine MHC-I with CTL epitopes

890	Interaction studie	s with Bol A-N·O	0901 and
		L epitopes	0501 4114
891	MHC-I	CTL epitopes	Binding energy (Kcal/mol)
892		LVLVGSLSY	-5.6
		FLYKRDLPY	-6.7
893		WMVFFIVVY	-6.4
		IAFCIILYY	-5.1
894	BoLA-N:00901	KMGHLTIYY	-6.6
0,54		STIAMGLVY	-5.6
		SSFNILLSY	-6.6
895		YSLLFFYLY	-6.3
		VVFDYSVKY	-5.6
896			
	Interaction stud	ies with BoLA-N:	00002 and CTI
897	interaction stud	epitopes	
	MHC-I		Di di di
898	WITC-1	CTL epitope	energy
898		115	-
898		LVLVGSLSY	energy
		115	energy (Kcal/mol)
899		LVLVGSLSY FLYKRDLPY WMVFFIVVY	energy (Kcal/mol) -6
		LVLVGSLSY FLYKRDLPY	energy (Kcal/mol) -6 -6.8
899		LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8
899		LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6
899 900	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVI SHY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6
899 900 901	BoLA-N:00902	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVI SHY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6
899 900	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVLSHY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6 (-6.1 -6.3
 899 900 901 902 	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVLSHY ILFTISLHY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6 -6.1 -6.3 (-6
899 900 901	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVLSHY ILFTISLHY KMGHLTIYY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6 -6.1 -6.3 (-6.3 (-6 -6 -4.4
 899 900 901 902 	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVLSHY ILFTISLHY KMGHLTIYY STIAMGLVY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6 (-6.1 -6.3 (-6 -6 -4.4 -5.6
899 900 901 902	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVLSHY ILFTISLHY KMGHLTIYY STIAMGLVY SSFNILLSY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6 -6.1 -6.3 (-6.1 -6.3 (-6 -4.4 -5.6 -5.9
 899 900 901 902 903 	10	LVLVGSLSY FLYKRDLPY WMVFFIVVY IAFCIILYY ILLTFNHLY KLVNLILLY MVIAVLSHY ILFTISLHY KMGHLTIYY STIAMGLVY SSFNILLSY YSLLFFYLY	energy (Kcal/mol) -6 -6.8 -6.1 -7.2 -4.8 -5.6 -6.1 -6.3 (-6.3 (-6 -4.4 -5.6 -5.9 (-5.5

Figure 01.TIF







B.





Figure 03.TIF





Figure 05.TIF



Figure 5.







Figure 06.TIF

